

OSTEOMYELITIS IN THE DIABETIC FOOT

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INTRODUCTION

Osteomyelitis is potentially one of the most serious problems seen in the diabetic foot. In a classic review, Waldvogel¹ determined that approximately one-third of all patients with osteomyelitis were diabetic. Certainly, the podiatrist will frequently be confronted with this pathologic process in the diabetic foot. One must be able to properly identify the presence of osteomyelitis, differentiating it from diabetic osteoarthropathy, and then develop a proper treatment plan. In many areas, this diagnostic and therapeutic process is still very controversial and one must have a working knowledge of both the pathophysiology of diabetes mellitus and osteomyelitis as a basis for rational decisions in the treatment of this dreaded combination.

DEFINITION

Osteomyelitis can be defined as an inflammation of bone marrow which most commonly presents as a pyogenic infection of bone marrow and/or bone.² The term osteomyelitis has frequently been used interchangeably with the term osteitis which technically describes the inflammation of bone. However, one must be aware that osteitis like osteomyelitis can represent an inflammatory process due to pyogenic infection of bone. Buckholz³ attempted to clarify this confusion by describing osteomyelitis as "an infection involving the marrow cavity or growth plate" and osteitis as "an infection of bone tissue that does not penetrate the medullary cavity or involve the growth plate." For practical purposes, this differentiation is only important in certain classification schemes with related treatment suggestions.

Pathophysiology

Many frequently misunderstood terms are used when attempting to describe the pathogenic process and findings in cases of osteomyelitis. Terms such as sequestrum, involucrum, and cloacae are distinctly defined osseous changes. When properly understood these findings can be used to obtain a more thorough understanding of the pathogenesis of the disease.

Osteomyelitis occurs when factors are present which favor the localization of bacteria. Obviously, circulatory compromise of diabetic angiopathy can create an environment suitable for the proliferation of bacteria via small areas of gangrene or skin ulceration with associated necrotic tissue. Other factors in the diabetic foot can also contribute to such localization, including sensory or autonomic changes frequently seen with diabetic neuropathy. A decrease in sensation may be associated with repetitive mechanical, thermal, or chemical trauma to the foot resulting in local tissue necrosis or ulceration. This may even occur without local vascular insufficiency. Osteoarthropathy may also be associated with sensory dysfunction creating new pressure areas for the repetitive trauma. Further, autonomic dysfunction may impair the vascular response to local tissue damage allowing pH changes which will be favorable to bacterial proliferation.⁴ Thus, both angiopathy and neuropathy together or individually can contribute to the localization of bacteria and eventual development of osteomyelitis. The adage that vascular insufficiency is the only cause for diabetic osteomyelitis has been disproved.

Once localization of bacteria has occurred, further destructive changes may become appar-

ent. The bacteria and any associated inflammatory reaction and pH change each contribute to the breakdown of trabecular and removal of matrix with calcium deposits. Therefore, the earliest radiographic change suggesting osteomyelitis is a loss of bone density or radiolucency.

As the infection proliferates, it spreads to neighboring osseous structures through the Haversian and Volkmann channels. This leads to the destruction of vascular channels and to additional necrosis and osteocyte death. As bone dies, it becomes sclerotic and will appear as such on radiographs. Large segments of devascularized dead bone can become separated to form sequestra (isolation of dead bone from living bone) (Fig. 1).⁴

Radiographic identification of bone sequestration is highly suggestive of chronic osteomyelitis. However, in the diabetic foot it must be carefully differentiated from sequestrum associated with osteoarthropathy.

If allowed to progress, the infection will eventually reach the subperiosteal area (Fig. 2). When the periosteum is elevated from the cortex by the suppurative process exuberant growth of new bone (involucrum) will be initiated (Fig. 3). The radiographic identification of subperiosteal involucrum formation is highly indicative of osteomyelitis.

Finally, a cloacae may form at the bone-periosteal interface and this represents an opening in the region for the extrusion sequestered bone (Fig. 4).⁵

Identification and an understanding of the pathophysiology and etiologic causes of osteomyelitis are paramount in its proper treatment and in the prevention of recurrence.

Classification

Osteomyelitis has been classified in numerous ways. Each classification has offered some assistance in understanding the disease process, but no system has yet been developed nor accepted which adequately describes the disease process and its relation to preferred treatment. Perhaps this is why so many classification and treatment systems exist.

Traditionally, osteomyelitis has been classified as an acute, subacute, or chronic infection based upon the clinical course of the disease and

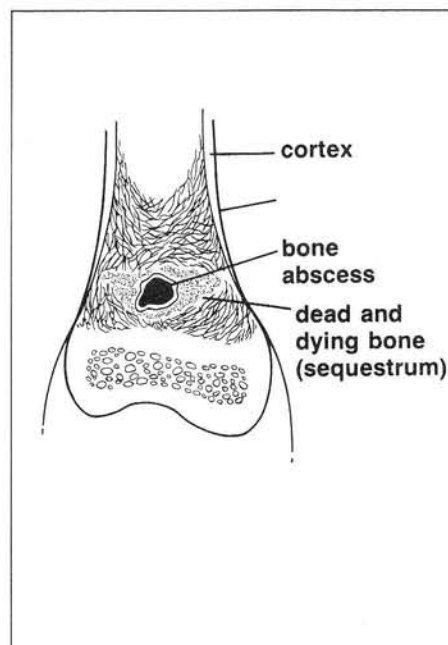


Fig. 1. Sequestrum: devascularized dead bone.

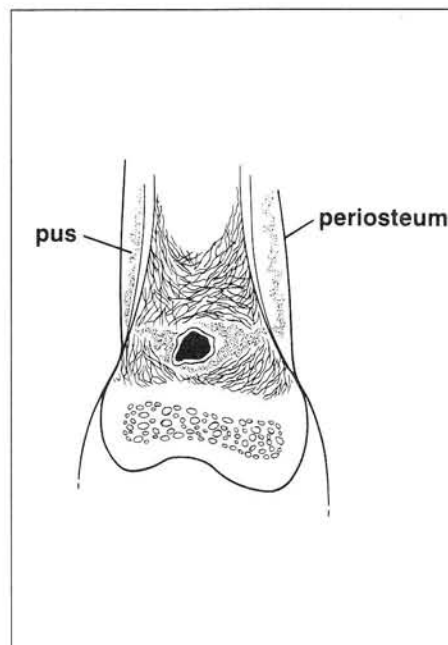


Fig. 2. Infection eventually reaches subperiosteal area. If close and intracapsular, then septic arthritis occurs.

the histological findings.⁶ This classification has always suffered from debate over the strict definitions for acute, subacute, and chronic osteomyelitis. In 1970, Waldvogel⁷ described a classification based loosely upon the pathogenesis of the disease. He divided osteomyelitis into three types of acute osteomyelitis: 1) hematogenous osteomyeli-

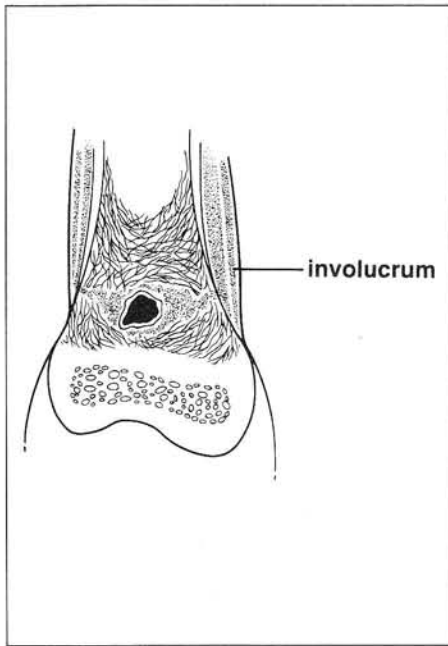


Fig. 3. Involucrum: new bone formation.

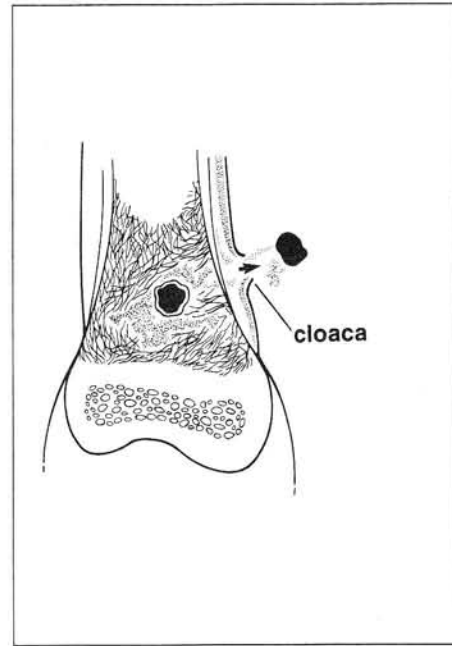


Fig. 4. Cloaca: Opening for extrusion of sequestrum and necrotic products.

tis, 2) osteomyelitis secondary to a contiguous focus of infection, 3) osteomyelitis associated with vascular insufficiency, and chronic osteomyelitis (Table 1). This classification may permit some suggestion of possible causative organisms and recommended therapy. However, there are certainly too many variables which this classification does not address (e.g., what is the contiguous focus of infection, an ulcer?, a puncture wound?) and, thus it is of limited usefulness. This is particularly true in the diabetic patient where osteomyelitis may be due to contiguous spread, vascular insufficiency, or both. Thus, Waldvogel's classification fails to consider the quality of the host, the anatomic nature of the disease, treatment factors or prognostic factors.

Subsequently, Cierny and Mader⁸ described a somewhat useful clinical staging system for osteomyelitis in adults using both an anatomic classification (stage I: medullary; stage II: superficial; stage III: localized, and stage IV: diffuse) and a physiologic classification (A-host: good systemic defenses, good local vascularity; B-host: systemic compromise, local compromise; C-host: not a surgical candidate, treatment worse than disease) (Table 2). Thus, by taking the three anatomic types and mixing them with each physiologic type, twelve different clinical stages or types of osteomyelitis are described. Treatment is then

TABLE 1

WALDVOGEL CLASSIFICATION SYSTEM

- Acute Osteomyelitis
 - Hematogenous osteomyelitis
 - Contiguous focus osteomyelitis
 - Osteomyelitis associated with vascular insufficiency
- Chronic Osteomyelitis

varied based upon these twelve stages of osteomyelitis. This staging system has been used at the University of Texas Medical Branch, and a more recent report described the results of treatment in 189 patients.⁹ Although not yet widely accepted, this system is a solid attempt at a practical classification scheme.

Buckholz⁵ described yet another classification system based upon the strict definitions of osteitis and osteomyelitis. He described seven types of bone infection: 1) wound induced, 2) mechanogenic infection, 3) physal osteomyelitis, 4) ischemic limb disease, 5) combinations of 1-4, 6) osteitis with septic arthritis, and 7) chronic

osteitis/osteomyelitis (Table 3). The classification system is rather confusing and as Buckholz states, "Unfortunately, familiarity with all seven types is not common to medical or surgical specialties. Each specialty encounters certain types of infection, and as a result, misunderstanding may arise between surgeon and clinician as to correct treatment." The classification may become useful if it becomes more widely understood and accepted.

Although no specific classification exists that is universally accepted, certain principles are common to the disease and will be discussed in relation to osteomyelitis in the diabetic foot. Any classification system may be helpful if it allows more accurate description and communication between the various medical disciplines involved in a patient's care.

DIAGNOSIS

The diagnosis of osteomyelitis in the diabetic foot can be a challenging and perplexing problem. Indeed, the diagnosis is frequently more difficult

than the treatment itself. The clinician must always be alert to the possibility of osteomyelitis. Frequently, the disease is overlooked if the cardinal signs of infection are absent, if leukocytosis is absent, or if soft tissue cultures are misleading or negative. The diagnosis of osteomyelitis depends on an accurate evaluation of clinical findings combined as necessary with radiographs, nuclear medicine studies, multiplanar imaging, laboratory studies, and bone biopsies and cultures.

Clinical Evaluation

Clinical evaluation includes a thorough history and physical examination. Examination for systemic and local signs of infection must be done. Any examination where infection is a possibility should begin with the taking of the patient's vital signs including body temperature. Locally, the cardinal signs which one should evaluate and monitor are edema, erythema, and increased temperature in the area. Most importantly, one must remember in diabetes that immunopathy is frequently present impairing the patient's response to inflammation and/or infection.¹⁰

Radiographic Examination

As stated earlier, classic radiographic changes associated with osteomyelitis include initial radiolucency followed by sclerosis, sequestrum formation, and involucrum formation. It is commonly believed that 10-14 days must pass before the initial radiolucency associated with osteomyelitis can be visualized radiographically. However, with careful observation, particularly when baseline or previous radiographs have been taken, radiolucency may be visualized within 5-7 days after the onset of infection.

TABLE 2

CIERNY-MADER CLASSIFICATION SYSTEM

Anatomic Stage

- Stage 1 - Medullary osteomyelitis
- Stage 2 - Superficial osteomyelitis
- Stage 3 - Localized osteomyelitis
- Stage 4 - Diffuse osteomyelitis

Physiologic Stage

- A Host - Normal host
- B Host - Systemic compromise (Bs)*
Local compromise (Bl)*
- C Host - Treatment worse than disease

*Systemic and Local Factors - B Host

SYSTEMIC FACTORS (Bs)	LOCAL FACTORS (Bl)
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- | | |
|---------------------|---------------------------|
| Malnutrition | Chronic lymphedema |
| Renal/Liver failure | Venous stasis |
| Alcohol abuse | Major arterial compromise |

Immune deficiency	Arteritis
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- | | |
|-------------------|--------------------|
| Malignancy | Extensive scarring |
| DIABETES MELLITUS | Radiation fibrosis |

- Extremes of age
- Steroid therapy
- Tobacco abuse

TABLE 3

BUCKHOLZ CLASSIFICATION SYSTEM

- 1 - Wound Induced
- 2 - Mechanogenic Infection
- 3 - Physeal Osteomyelitis
- 4 - Ischemic Limb Disease
- 5 - Combinations of 1-4
- 6 - Osteitis with Septic Arthritis
- 7 - Chronic Osteitis/Osteomyelitis

Nuclear Medicine Studies

Nuclear medicine studies may be particularly helpful in the diagnosis of osteomyelitis. In some cases, they may be particularly helpful in differentiating osteomyelitis from osteoarthropathy. Technetium-99m methylene diphosphonate (Tc-99), gallium-67 citrate (Ga-67), and indium-111 oxine (In-111) are frequently used as imaging agents to aid in the diagnosis of osteomyelitis. Each has specific properties, advantages, and limitations.

The technetium-99 bone scan is the most widely used study to document the evidence of osteomyelitis. However, Tc-99 is known for its sensitivity, but not for its specificity. It is very important to utilize a three- or even four-phase bone scan to differentiate infection of the soft tissues around bone from infection within bone. The three-phase bone scan is composed of a radionuclide angiogram (first phase) - demonstrating the dynamic blood flow to the area, an immediate post-injection blood pool image (second phase) - representing relative vascular flow, and a 2 to 4 hour delayed image (third phase) - reflecting skeletal uptake. Evidence suggests that a fourth phase, another delayed image taken 24 hours post-injection, may be helpful.¹¹

The first phase consists of several images taken in rapid sequence 1 to 2 seconds apart as the isotope approaches the extremity. This affords a dynamic visualization of blood flow to the extremity and functions much like an angiogram. The second phase is also a vascular phase and is taken only minutes after injection of the isotope. After the first several minutes, an equilibrium is reached throughout the blood volume and a series of images can be taken to demonstrate the "blood pool." This term accurately describes the quantity of blood "pooling" or present in the capillary beds and veins. Thus, the first two phases demonstrate the vascularity to the region and will be "hot" whether the infectious process is soft tissue or bone.

The delayed image phases (third and fourth phases) take advantage of the Tc-99 as a "bone-seeking" isotope or an isotope which demonstrates the amount of osteoblastic activity. By taking an image 2 to 4 hours after injection, one has allowed osteoblasts to use the isotope in the production of new bone, whether for repair or main-

tenance. One has also allowed the excretion of most of the unused isotope since the half-life of Tc-99 is about 6 hours. The greater the delay, the more the bone activity and the less the soft tissue activity. The fourth phase may actually prove to be the most valuable although dropping activity may make imaging more difficult. In the diabetic with severe vascular disease, the isotope may localize in bone very slowly and be excreted very slowly as well. An image 5-24 hours after injection may be more helpful in diagnosing osteomyelitis in such instances.¹²

The best evaluation of an osteomyelitis process comes from the evaluation of all of the phases simultaneously. The earlier phases will be "hot" in the face of soft tissue and/or bone infection. However, if a soft tissue infection alone is present the third and fourth phases should demonstrate comparatively less activity and any activity present should be more diffuse. A Tc-99 bone scan that does not demonstrate appreciable activity in the third and fourth phases significantly decreases the probability of osteomyelitis. If the third and fourth phases seem to demonstrate similar or greater activity with discrete focal uptake, then osteomyelitis may be suspected.

Difficulty arises in the diabetic with osteoarthropathy in that the bone scan may be "hot" in all phases due to the bone activity associated with this hyperemic and pathogenic process. One must recognize this limitation, but with skill and experience one can still utilize Tc-99 bone scans and specific diagnostic patterns to occasionally establish the presence or absence of osteomyelitis even with concomitant osteoarthropathy.

Gallium-67 scanning is used predominantly for the detection of acute inflammation or infection since the isotope binds to white blood cells and plasma proteins. Ga-67 alone is not indicated for the detection of osteomyelitis in the diabetic foot. The concurrent use of Tc-99 bone scans and Ga-67 may have some practical use in the diagnosis of osteomyelitis. This approach uses a Tc-99 bone scan and a Ga-67 obtained 24 to 72 hours later. Acute osteomyelitis can be more intensely "hot" on Ga-67 scanning compared to simultaneous Tc-99 bone scans. On the other hand, if gallium uptake is less than that of technetium, an osteomyelitic process is unlikely. In more chronic cases of osteomyelitis the Ga-67 scan theoretically should be negative while the Tc-99 bone scan

should be "hot." However, in practice the Ga-67 scan demonstrates variable activity in cases of chronic osteomyelitis.¹³

Many investigators have reported that Ga-67 scanning may be useful in following the success of therapy oriented at the sterilization of osteomyelitis.¹⁴⁻¹⁷ Tc-99 bone scans may be positive for months or years due to continued bone remodeling, while Ga-67 scans will generally become negative as the infection is arrested. However, the use of Ga-67 scans for this purpose is still controversial and clinical studies have reported positive Ga-67 scans following the complete clinical resolution of the osteomyelitis.¹⁸

The Indium-111 white blood cell scan may be the most helpful in the diabetic foot. A comparative report concluded that In-111 scanning is superior to combined bone and gallium imaging in patients with musculoskeletal sepsis.¹⁹ Early evidence also suggests that it may be of use in differentiating osteomyelitis from osteoarthritis. To perform this study neutrophils isolated from blood taken from the patient are labelled with In-111. The tagged In-111 white blood cells are then injected back into the patient and the scan performed approximately 18-24 hours later. The neutrophils localize in the inflammatory area, whether bone or soft tissue. The scan is both highly sensitive and highly specific for acute soft tissue and osseous infection. Thus, it can be used to differentiate acute osteomyelitis from osteoarthritis in the diabetic foot. The In-111 scan should be negative in the osteoarthritis and "hot" (positive) in the presence of acute osteomyelitis. Indium-111 scanning may not be beneficial in chronic osteomyelitis due to a predominately lymphocytic pattern that will not cause localization of the In-111 labelled neutrophils.²⁰

CT/MR Imaging

Computed tomography (CT) and magnetic resonance imaging (MRI) can be useful in the diagnosis and management of osteomyelitis, particularly when conventional radiographs and nuclear medicine studies are equivocal.

Cortical bone destruction (i.e., sequestrum), periosteal proliferation (i.e., involucrum), and soft tissue extension can readily be visualized on CT.

CT offers cross-sectional visualization in the frontal and transverse planes. Sagittal plane views must be indirectly reformatted from either frontal or transverse plane views.²¹ CT is generally inferior to MRI in the evaluation of medullary bone, although the presence of gas within the medullary cavity as detected by CT has been described as a diagnostic sign of osteomyelitis.²² This phenomenon has been attributed to the infiltration of bone by organisms from the surrounding soft tissues, resulting in infection of the medullary cavity. The causative organisms have not been identified in reported cases of intraosseous gas, and osseous abnormalities have been uniformly absent on standard radiographs.

MRI has high sensitivity for inflammatory processes in either medullary bone or soft tissue. MRI like CT can provide cross-sectional imaging. Although clinical experience utilizing this modality is limited, preliminary evidence suggests that MRI has a detection sensitivity for musculoskeletal infection that approaches that of radionuclide studies.²³ However, due to its cost, this modality will remain an adjunct to be used for localizing osteomyelitis in difficult cases.

Laboratory Studies

Laboratory studies are more useful for following the treatment of osteomyelitis than in diagnosing it. The complete blood count (CBC) may be normal as previously mentioned. The Westergren erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP) may be positive, but are only a general indicator of inflammation. They may also be positive in osteoarthritis.

Most of the laboratory studies are helpful in following the therapeutic course of osteomyelitis. If the CBC demonstrated an elevated white blood cell count and differential shift, it should demonstrate a marked decrease in the white blood cell count and band cell count after initiation of appropriate treatment. The ESR and CRP have also been reported to be helpful in monitoring the clinical improvement in the treatment of osteomyelitis, particularly in pseudomonas osteomyelitis. If the ESR was elevated, it should decrease with treatment, and if the CRP was positive it should become negative with treatment.^{24,25}

Bone Biopsy and Culture

Bone biopsies and cultures are the best and only definitive evidence of osteomyelitis. They are also the only definitive method to differentiate osteomyelitis from osteoarthropathy. The "key" to the appropriate treatment and antibiotic selection in any infection, including osteomyelitis, is to obtain reliable cultures. Soft tissue cultures and those obtained from sinus tracts and overlying ulcers are notoriously poor cultures and very unreliable.^{26,27} Bone cultures from the involved bone should be obtained whenever possible in cases of osteomyelitis. Care must be taken to obtain the cultures without passing through infected soft tissue and with the patient withdrawn from any antibiotics for at least 48 hours. Biopsies and cultures are best obtained through open exposure as blind biopsies with a needle or trephine can lead to false-negative cultures and pathology reports.

TREATMENT

Norden²⁸ experimentally demonstrated that osteomyelitis is difficult to induce, but once established is even more difficult to eradicate. The treatment of osteomyelitis is still controversial in many areas. However, it is now generally accepted that surgical excision of the osteomyelitic bone combined with intravenous antibiotics should be instituted whenever possible. Only in very early cases of acute osteomyelitis or in chronic hematogenous osteomyelitis should antibiotic therapy alone be considered.

Once osteomyelitis is diagnosed, its surgical excision should be planned. In the diabetic with profound vascular insufficiency appropriate evaluation must be undertaken to determine the potential for healing prior to the surgical procedure for the osteomyelitis. For the podiatric surgeon, this may involve referral to a vascular surgeon for evaluation and possible revascularization procedures. Once the area demonstrates satisfactory vascularity, the surgical procedure may be performed. If severe infection and necrosis occurs prior to such revascularization, then amputation may need to be considered at a more proximal level where the healing potential has been determined to be satisfactory.

Whenever possible all of the osteomyelitic bone should be excised. Surgical approaches can be through a previous area of infection or through clean incisional areas. All necrotic bone, soft tissues, and devascularized structures should be excised and wound revision performed to accommodate immediate closure or closure at a later date. Debate continues as to how much bone should be excised. Certainly, all infected bone and a small portion of apparently uninvolved bone should be removed. The uninvolved bone may be sent for separate microscopic evaluation. If the infected bone exhibits microscopic evidence of osteomyelitis and the uninvolved bone is reported as noninfected, then one has documented the excision of the diseased bone.

Further debate continues as to the advantages and disadvantages of disarticulation versus transcortical amputation in areas of joint involvement. Disarticulation maintains the subchondral bone and cartilage as a natural barrier to the spread of the infection into the remaining osseous structure. Transcortical amputation permits the microorganisms access to the Haversian and Volkmann channels but excises the cartilage which will certainly become necrotic if left for several days in the open wound. A possible compromise is to perform a disarticulation and then gently curette the cartilage from the remaining portion of bone. The subchondral bone will provide a better barrier than is present with transcortical amputation.

Once surgical excision of osteomyelitic bone has been performed a method of wound care must be chosen. Primary closure with the use of antibiotic-impregnated beads, primary closure with the use of a closed suction irrigation system, and open packing are all acceptable alternatives. If open packing is chosen, the wound may be closed after subsequent wound cultures have documented the eradication of the infection.

All treatment regimens should include the use of intravenous antibiotics. The appropriate antibiotic should be chosen and oriented toward the pathogenic organism(s) identified by reliable culture.²⁹ Today, appropriate antibiotic therapy is considered to be 6 weeks of parenteral antibiotic administration. Frequent relapse and chronic osteomyelitis can be anticipated with short-term or oral antibiotic therapy.³⁰ The patient may be

discharged and continued on intravenous antibiotics at home if the services of a home health care team or nurse are available. This approach lowers cost significantly and drastically improves the patient's overall acceptance of lengthy therapy.³¹

Another possible adjunctive mode of therapy is hyperbaric oxygen (HBO). The mechanisms of action of HBO are not entirely understood. In vitro studies have indicated that increased atmospheric pressures of oxygen inhibit the growth of both aerobic and anaerobic bacteria.^{32,33} However, the partial pressure of oxygen necessary to show this activity in vitro is unattainable in vivo. When hyperbaric oxygen was used under conditions applicable to human tissue, no inhibition of aerobic organisms was noted.³⁴

Despite this evidence, several human clinical trials have reported HBO to be an effective treatment modality. The first of these in vivo trials was reported by Slack et al.³⁵ in 1965. They noted five patients with recurrent osteomyelitis who responded to HBO alone. Subsequent studies have generally combined HBO with sequestrectomy and antibiotics. Their reported success rates vary from 70%-85%, but lack adequate follow-up and comparative trials.

Realizing the discrepancy between the apparently promising clinical results and the inability to get theoretically anti-bacterial partial oxygen pressures in vivo, several hypotheses have been offered as to the actual mechanisms of action on HBO. The most plausible explanation appears to be the beneficial effect of HBO on tissue hypoxia and phagocytic function.³⁶ This hypothesis is supported by the fact that studies have shown local extremity hyperbaric chambers to be ineffective. Only full body hyperbaric chambers can increase systemic oxygen pressures sufficiently to increase the polymorphonuclear leukocyte's killing effect of bacteria.³⁴

Apparently then, hyperbaric oxygen when administered in full body hyperbaric chambers may be beneficial in the treatment of osteomyelitis. The availability of treatment chambers and the time needed for treatment have limited the size of clinical studies. Good clinical trials with comparative studies need to be performed to obtain more definitive evidence as to the efficacy of hyperbaric oxygen.

SUMMARY

Osteomyelitis in the diabetic foot is one of the more challenging diagnostic and treatment problems encountered by the podiatric physician and surgeon. Differentiation of the disease process from diabetic osteoarthropathy can be particularly difficult. An appreciation of the pathogenesis of diabetes and osteomyelitis is critical in aiding diagnosis and in the development of an appropriate treatment plan.

REFERENCES

1. Waldvogel FA, Medoff G, Swartz MN: Osteomyelitis: a review of clinical features, therapeutic considerations and unusual aspects. III. *N England J Med* 282:316-322, 1970.
2. Downey MS, Jimenez AL: Osteomyelitis. In McGlamry ED (ed): *Doctors Hospital Podiatric Education and Research Institute Fifteenth Annual Surgical Seminar Syllabus*. Atlanta, Doctors Hospital Podiatric Education and Research Institute, 1986, pp 83-88.
3. Buckholz JM: The surgical management of osteomyelitis: with special reference to a surgical classification. *J Foot Surg* (supplement) 26:S17-S24, 1987.
4. Joseph WS, LeFrock JL: The pathogenesis of diabetic foot infections - immunopathy, angiopathy, and neuropathy. *J Foot Surg* (supplement) 26:S7-S11, 1987.
5. Kehr LE, Zulli LP, McCarthy DJ: Radiographic factors in osteomyelitis. *J Am Podiatry Assoc* 67:716-732, 1977.
6. Roth RD, Pressman M: Clinical diagnosis of osteomyelitis. *J Am Podiatry Assoc* 67:709-715, 1977.
7. Waldvogel FA, Medoff G, Swartz MN: Osteomyelitis: a review of clinical features, therapeutic considerations and unusual aspects. I. *N England J Med* 282:198-206, 1970.
8. Cierny G, Mader JT: Adult chronic osteomyelitis. *Orthopedics* 7:1557-1564, 1984.
9. Cierny G, Mader JT, Penninck JJ: A clinical staging system for adult osteomyelitis. *Contemp Orthop* 10:17-37, 1985.
10. Banks AS: Diagnosis of infection. In McGlamry ED (ed): *Doctors Hospital Podiatric Education and Research Institute Fifteenth Annual Surgical Seminar*. Atlanta, Doctors Hospital Podiatric Education and Research Institute, 1986, pp 62-67.
11. Alazraki N, Dries D, Datz F, Lawrence P, Greenberg E, Taylor A: Value of a 24-hour image (four-phase bone scan) in assessing osteomyelitis in patients with peripheral vascular disease. *J Nucl Med* 26:711-717, 1985.
12. Hartshorne MF, Peters V: Nuclear medicine applications for the diabetic foot. *Clinics in Podiatry* 4:361-375, 1987.
13. Rosenthal L, Kloiber R, Damteu B, et al: Sequential use of radiophosphate and radiogallium imaging in the differential diagnosis of bone, joint and soft tissue infection: quantitative analysis. *Diagn Imaging* 51:249-258, 1982.

14. Deysine M, Rafkin H, Teicher I, Silver L, Robinson R, Manly J, Aufses AH: Diagnosis of chronic and postoperative osteomyelitis with gallium 67 citrate scans. *Am J Surg* 129:632-635, 1975.
15. Graham GD, Lundy MM, Frederick RJ, Berger DE, O'Brien AW, Brown TJ: Predicting the cure of osteomyelitis under treatment: concise communication. *J Nucl Med* 24:110-113, 1983.
16. Graham GD, Lundy MM, Moreno AJ, Frederick RJ: The role of 99mTc MDP and 67Ga citrate in predicting the cure of osteomyelitis. *Clin Nucl Med* 8:344-346, 1983.
17. Alazraki N, Fierer J, Resnick D: Chronic osteomyelitis: monitoring by 99mTc phosphate and 67Ga citrate imaging. *AJR* 145:767-771, 1985.
18. Kolyvas E, Rosenthal L, Ahronheim G, et al: Serial 67Ga-citrate imaging during treatment of acute osteomyelitis in children. *Clin Nucl Med* 3:461-466, 1978.
19. Merkel KD, Brown ML, Dewanjee MK, et al: Comparison of indium-labeled-leukocyte imaging with sequential technetium-gallium scanning in the diagnosis of low-grade musculoskeletal sepsis. *J Bone Joint Surg* 67A:467-476, 1985.
20. Sheikh WA, Sfakianakis GN, Mnaymneh W, Hourani M, Heal A, Duncan RC, Burnett A, Ashkar FS, Serafini AN: Subacute and chronic bone infections: diagnosis using In-111, Ga-67 and Tc-99m MDP bone scintigraphy, and radiography. *Radiology* 155:501-506, 1985.
21. Wing VW, Jeffrey RB, Federle MP, et al: Chronic osteomyelitis examined by CT. *Radiology* 154:171-174, 1985.
22. Ram PC, Martinez S, Korobkin M, et al: CT detection of intraosseous gas: a new sign of osteomyelitis. *AJR* 137:721-723, 1981.
23. Modic M, Pflanze W, Helhobek G: Magnetic resonance imaging of musculoskeletal infections. *Radiol Clin North Am* 24:247-259, 1986.
24. Peltola H, Vahvanen V, Aalto K: Fever, C-reactive protein and erythrocyte sedimentation rate in monitoring recovery from septic arthritis: a preliminary study. *Journal Pediatric Orthopedics* 4:170-174, 1984.
25. Crosby LA, Powell DA: The potential value of the sedimentation rate in monitoring treatment outcome in puncture-wound related pseudomonal osteomyelitis. *Clin Orthop* 188:168-172, 1984.
26. Mackowiak PA, Jones SR, Smith JW: Diagnostic value of sinus tract cultures in chronic osteomyelitis. *JAMA* 239:2772-2775, 1978.
27. Sapico FL, Witte JL, Canawati HN, Montgomerie JZ, Bessman AN: The infected foot of the diabetic patient: quantitative microbiology and analysis of clinical features. *Rev Infect Dis* 6:S171-S176, 1984.
28. Norden C: Experimental osteomyelitis. I. A description of the model. *J Infect Dis* 122:410-418, 1970.
29. Downey MS: Principles of antibiotic therapy. In McGlamry ED (ed): *Doctors Hospital Podiatric Education and Research Institute Fifteenth Annual Surgical Seminar Syllabus* Atlanta, Doctors Hospital Podiatric Education and Research Institute, 1986, pp 68-82.
30. Jacob LS: Pharmacological therapy in the treatment of osteomyelitis. *J Am Podiatry Assoc* 67:706-708, 1977.
31. Grizzard MB: Home intravenous antibiotic therapy: a practical management approach for the 1980s. *Antibiotic Therapy* 78:187-195, 1985.
32. Hopkinson WI, Tower AG: Effects of hyperbaric oxygen on some common pathogenic bacteria. *Lancet* 2:1361-1363, 1963.
33. McAllister TA, Stark JM, Norman JN, Ross RM: Inhibitory effects of hyperbaric oxygen on bacteria and fungi. *Lancet* 2:1040-1042, 1963.
34. Mader JT, Brown GL, Guckian JC, Wells CH, Reinartz JA: A mechanism for the amelioration by hyperbaric oxygen of experimental staphylococcal osteomyelitis in rabbits. *J Infect Dis* 142:915-922, 1980.
35. Slack WK, Thomas DA, Perrins D: Hyperbaric oxygenation in chronic osteomyelitis. *Lancet* 1:1093-1094, 1965.
36. Braun TI, Lorber B: Chronic osteomyelitis. In Schlossberg D (ed): *Orthopedic Infection*. New York, Springer-Verlag, 1988, pp 9-20.